

Poly sulfide Quantification by Dynamic Headspace and Gas Chromatography-Sulfur Chemiluminescence Detection Extraction method: Sample (250 mL) was poured into a 500-mL flat-bottomed flask fitted with a sterilized Druschel head. The flask was placed in a thermostatic bath maintained at 30°C. A Tenax cartridge (90 mg, 25–30 mesh; Chrompack, Sint-Katelijne-Waver, Belgium) was fitted to the gas vent branch of the Druschel head, and another was attached to the purge unit. Volatiles were purged into the Tenax cartridge for 10 min with a 30-mL/min nitrogen flow. The Tenax cartridge was then dried using an inverted 15-mL/min nitrogen current for 3 min and transferred to the gas chromatographic (GC) unit (TCT/Pt 4001, Chrompack, Antwerp, Belgium) to determine the total sulfur content.

Accelerated Aging of Bottled Beers Spiked with Copper II ($\text{CuCl}_2 \cdot 2\text{H}_2\text{O}$, 10 mM) and Ascorbic Acid (10 mM) were injected into bottled commercial Lager beers. The bottles were then closed with a silicone top (No. 5, Vel, Leuven, Belgium), crown sealed, and aged at 40°C for five days in a dark room.

A control beer was produced with 50 ppm of SO_2 , or 25 ppm of ascorbic acid were added during mashing in. Mashings were performed in a 30-L jacketed kettle (Biosster U, B. Brauun, Vilvoorde, Belgium) with 8.5 kg malt flour and 21.25 L of water (Millipore, concentrate 35 ppm $\text{CaSO}_4 \cdot 2\text{H}_2\text{O}$, 10 ppm $\text{MgCl}_2 \cdot 6\text{H}_2\text{O}$, and 30 ppm NaCl). The temperature regime applied was 50°C for 30 min, to 63°C at 1.3 degrees C per minute, 63°C for 30 min, to 72°C at 0.6 degrees C per minute, and 72°C for 30 min. The temperature was then raised to 80°C at 0.6 degrees C per minute, and the wort was filtered with a "2001 filter" (Meura, Louvain-la-Neuve, Belgium). The clarity of the filtered wort was adjusted to 12°P with mashed water and boiled for 75 min. After 20 min of clarification, the tube was eliminated by filtration, and 0.3 ppm of ZnCl_2 was added to the wort. Fermentation was conducted in 3-L EBC tubes with an initial yeast (Saccharomyces cerevisiae, pitching rate: 7.5×10^6 cells/ml liter) at 20°C for seven days and 7°C for seven days.

EXPERIMENTAL

Effect of the Reducing Power of a Beer on Dimethyltrisulfide

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In the main precipitation of dimethyltinustannide in aged lead, tin tetrabutyltinate may play a role in this reaction. It is known that tin tetrabutyltinate is a strong reducing agent, and it is also known that tin tetrabutyltinate can reduce the tin(IV) in tin tetrabutyltinate to tin(II).¹ The reduction of tin(IV) to tin(II) may be due to the presence of the tin(IV) in tin tetrabutyltinate, which is reduced by the tin tetrabutyltinate. The reduction of tin(IV) to tin(II) may be due to the presence of the tin(IV) in tin tetrabutyltinate, which is reduced by the tin tetrabutyltinate.

NEWOSCAN

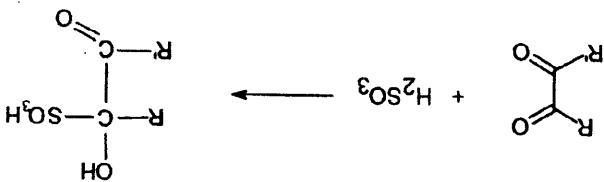
Palabras claves: Envejecimiento de cerveza, Mezcal, Mezcalero, Mezcalizante, Mezcalizante de dimetilpropionilaldehido, Mezcalizante de dimetilpropionilalcano, Mezcalizante de dimetilpropionilalcano con cobre, Estos tipos de enzima conducen a mayores o menores niveles de dimetilpropionilalcano.

Hydroxyl radicals, produced either with copper (II) and ascorbic acid or iron (II) and hydrosigen peroxide, can favor decomposition of dimethyltrisulfide (DMTS) in hops as the postulated precursor of dimethylcysteine sulfoxide (DMCS) in aged beers (6), but Gijss and coworkers (3) have recently proposed two additional DMTS sources in aged beers: 3-methyltrisulfone and its reduced form, 3-methyltrisulfide. These results suggest that the rate of DMTS synthesis during aging is the best indicator of the rate of DMS synthesis during aging. Analogous to the well-known nonenzymatic, it was defined as the DMTS potential (2).

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TABLE I		Dimeethyltrisulfide Concentration in Aged Beers from the Control Production and the Production with Sulfite or Ascorbic Acid Added at 20 ppm of Ascorbic Acid					
Production		Results of Two Trials		Averagge		With 20 ppm of SO ₂	
Control		98		98		98	
With 20 ppm of Ascorbic Acid		104		99		332	
With 50 ppm of SO ₂		321		309		98	
With 50 ppm of SO ₂		98		98		98	
With 20 ppm of Ascorbic Acid		104		99		108	

Fig. 2. Reaction between sulfite and diketone or a reducing sugar.



amounts of DMTs in the aged beer similar to those found in the addition of 20 ppm ascorbic acid during mashing-in leads to impact of Ascorbic Acid

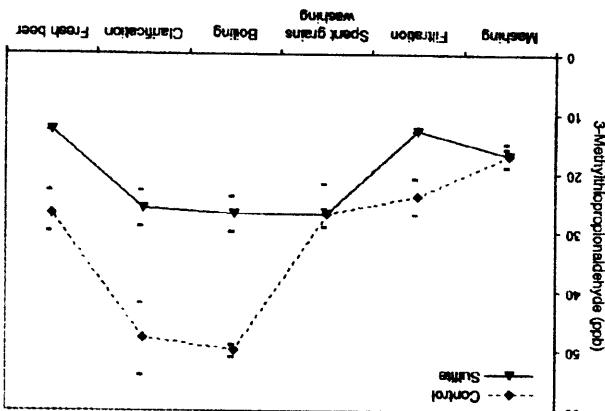
(Fig. 3b) during aging would then increase polysulfide synthesis. Therefore, we suggest that sulfites act here, through their capacity to bind 3-methylthiopropionaldehyde (Fig. 3a). This DMTs protein (Table I).

During fermentation, as expected, 3-methylthiopropionaldehyde reduced sulfur was released in the aged beer despite its low amounts of DMTs were measured in the aged beer extracted in the dark (Fig. 3b) during aging. The gradual release of methanethiol (Fig. 3c) is probably due to the formation of methanethiol (Fig. 3d), without losing the sulfite function.

Hence, a very low DMTs potential characterized our production inhibited (Fig. 2; mainly in the spent grains and in the kettle). Precursors in the wort, starch degradation of methionine is reduced due to the formation of sugars and diketone formed due to the conversion of sulfites, which mask the reducing power of sulfites, a lower volatility (7). In the presence of sulfites, more chemical reactions occur during boiling. Compared to 3- and 2-methylbutanal, evaporation losses are larger reactions. More chemical reactions occur during boiling. Methyliopropionaldehyde is produced in the 2001 filter by Mann control, as is the case for 3- and 2-methylbutanal (1). 3-Methylthiopropionaldehyde is added during the filtration (WCO) CP-SIIS CB (Chromopack), capillary open tubular column (0.32-mm x 0.32-mm, wall-coated open tubular column) (Fig. 1) shows the evolution of 3-methylthiopropionaldehyde DMTs during aging.

In the final product of as autoxidants during mashing-in of (4). Unknown, however, is the effect of sulfites on the 3-methylthiopropionaldehyde content of wort and subsequent production of DMTs during aging.

Fig. 1. Evolution of 3-methylthiopropionaldehyde during beer production with addition of 50 ppm sulfite at the mashing-in. Comparison with a control production.



The flavorable influence of sulfites on the *trans*-2-nonenal level in aged beer is well known. They act either by masking aldehydes

RESULTS AND DISCUSSION

Gases chromatography analysis (GC) equipped with flame ionization detector (model 5890, Hewlett-Packard, Golden, CO) was used. A gas chromatograph (model 3890, Hewlett-Packard, Kyoto, Japan), was used. An automatic sample injector (model 7673, Hewlett-Packard), a cold on-column injector, a flame ionization detector, and an integrator before transferring the extract to a chromatographic vial. Kudema Danish vessel. The organic phase was concentrated to 0.5 ml in a dichloromethane. The aqueous phase was concentrated to 0.5 ml in a and under stirring with, successively, 20, 15, and 15 ml of distilled water transferred to a glass tube. Each 25-ml sample was extracted in the dark (Fig. 1) shows the evolution of 3-methylthiopropionaldehyde DMTs during aging.

Liquid/liquid Extraction and Gas Chromatography-Flame Ionization Detection

3-Methylthiopropionaldehyde Quantification by Gas chromatography and GC analytical conditions. GC analyses were carried out on a 50-m x 0.32-mm, wall-coated open tubular column (0.32-mm x 0.32-mm, wall-coated open tubular column) (Fig. 1) shows the evolution of 3-methylthiopropionaldehyde DMTs during aging.

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LITERATURE

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Surprisingly, the reducing power of the medium proves not to be a key determinant of DMTS production during beer aging. Sulfites protect 3-methyltinopropionaldehyde against aging, but the aldehyde, thus making it available for methanethiol oxidation to 3-methyltinopropionic acid through their capacity to bind the sulfite. This makes it available for methanethiol release during aging. Addition of ascorbic acid during the mash-ing-in does not affect the DMTS potential. Despite its prooxidant properties, copper did strongly modify the flavor composition, but its ability to complex methanethiol or hydrogen sulfide is clearly the property to be taken into account to explain our results.

CONCLUSIONS

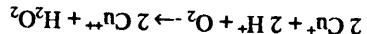
Influence of polysulfide synthesis. In our spiked beer, of course, copper did strongly modify the flavor composition, but its ability to complex methanethiol or hydrogen sulfide is clearly the property to be taken into account to explain our results.

Surprisingly, despite its prooxidant properties, copper II (10 mM CuCl₂) and ascorbic acid (10 mM), again, this experimentally inhibited DMTS synthesis (Table II). Again, this copper complex added to commercial beers spiked with copper II (10 mM CuCl₂) and ascorbic acid (10 mM).

To assess the impact of copper I on beer aging, we applied acetone-d₆ aging to commercial beers spiked with copper II (10

ethylene. To increase radical degradation of 3-methyltinopropionaldehyde to aqueous model medium, we could suspect that copper I in an aqueous model medium (the source of copper I) in an (4) with copper II and ascorbic acid (the source of copper I) in an aqueous model medium would increase radical degradation of 3-methyltinopropionaldehyde in aged beers.

Therefore, as previously shown by Lieberman and coworkers (*J. Am. Soc. Brew Chem* 57:29-33, 1999).



Copper I is known to generate hydroxyl radicals from hydrogen peroxide through the Fenton reaction:

Impact of Copper

appears not to be a key parameter for controlling the polysulfide content.

With control (Table I), the reducing power of the wort thus aged control (Table I). The reducing power of the polysulfide

Beer	Results of Two Trials		Dimethyltin sulfate (ppm)
	Average	Control	
With copper II (10 mM) and ascorbic acid (10 mM)	0	0	0
Without copper II (5 days at 40°C) Spiked or Not with Copper II and Ascorbic Acid Before Aging	47	49	50

TABLE II
Dimethyltin sulfate Concentration in Aged Beers (5 days at 40°C) Spiked or Not with Copper II and Ascorbic Acid Before Aging

Fig. 3. Hypothetical degradation of 3-methyltinopropionaldehyde.

